Tagging News

News from the ORI Cooperative Fish Tagging Project • Published July 2023 •Results from 2022 •Number 36







Helping people to care for our ocean

Gareth Jordaan

Welcome to the 36th edition of the Tagging News.



For nearly four decades the Tagging News has been communicating the results of the ORI Cooperative Fish Project (ORI-CFTP) Tagging to our members and has successfully promoted ethical angling while tracking the growth rates and movement patterns of many of the common species linefish caught along the southern African coast.

The ORI-CFTP was recently privileged to give an online

talk on the history and achievements of the tagging project throughout its 38-year duration (1984-2022), which was hosted by Leadership for Conservation in Africa (LCA). This talk was part of the Unlocking Nature series that LCA started in 2020 which allows conservationists and conservation companies to share their stories and adventures with audiences worldwide. This was a great opportunity for Dr Bruce Mann and I to share to both local and international audiences what the ORI-CFTP is all about, some of its achievements over the years and how it has helped contribute towards improving the conservation awareness and behaviour of marine recreational anglers. The talk was well received and led to some great insights and questions afterwards. If you missed the talk, not to worry, you can catch a recording of it here. Furthermore, the ORI-CFTP has been invited to join the 'World Volunteer Fishtag Summit 2023' which is going to be an online conference in August 2023 hosted by SUNTAG in Australia and will bring various cooperative fish tagging projects from around the world together to share, discuss and collaborate on the work they have been doing. These two platforms have played a crucial role in promoting global recognition for the ORI-CFTP. We are thrilled to have the opportunity to showcase the remarkable accomplishments of this project and to gain insights from others. The ORI-CFTP owes its current position and international collaboration prospects to the unwavering support and dedication of our tagging members. We deeply appreciate the tremendous effort each of you has contributed towards this project. Your contributions are invaluable, and we are truly grateful for your ongoing commitment! Thank you!

The year 2022 again yielded some fantastic results for the ORI-CFTP. Although the total number of new members was less (n = 249) than 2021 (most likely due to the implementation of a stricter application process by the

Tagging Officer), we still had a reasonably high number of tag releases (n = 11 525) and an increased recapture rate of 8.8%! Although some long-term tagging projects came to an end in 2021, others continued to thrive (see Table on page 15). Overall, the total number for fish tagged (n = 375 614) and recaptured (23 635; 6.3%) since the inception of the project is a truly exceptional effort!

In 2022 our top tagger was Nic de Kock (who has been a member of the tagging project since 1984) with 446 tag releases followed by, for the third year in a row, Mark Galpin with 268 fish tagged. Nic also had the greatest number of his tagged fish recaptured in 2022 with 29 (see Table on page 8). Just a reminder to all taggers that it is not about the number of fish you tag, but rather the way you catch, handle, tag and release your fish (see some helpful tips here) that is far more important. This results in a greater chance of your fish's survival and ultimately being recaptured. Furthermore, ensuring that your tagging data is accurately recorded and sent back to the Tagging Officer is of equal importance. Over the past year, the ORI-CFTP has also been emphasising the importance of the welfare of fish tagged through our new tagging video on capturing, landing, handling and releasing large sharks caught from the shore, as well as prohibiting our members tagging all ray species.

In this year's Tagging News, you can look forward to reading some great articles including a 21-year history of fish monitoring and tagging in the iSimangaliso Marine Protected Area (Page 4) and the interesting work that is being done by the Coega Harbour Tagging Project (Page 18). You can also read about the Acoustic Tracking Array Platform (ATAP) and how this cutting-edge project is helping to 'fill in the gaps' in our knowledge of the finer movement patterns we are not able to pick up from external dart tagging done by the ORI-CFTP (Page 24). Our focus species for this year is the catface rockcod, because the ORI team recently published a paper on the movement patterns of this enigmatic species using both the ORI-CFTP and ATAP data (Page 27).

For those of you on social media, please remember to give the **ORITag FB page** a 'like' and share it with your angling buddies. Please also like and share the new **@ori_tagging_project** Instagram page. We strongly encourage those of you who have not yet seen our **instructional tagging videos** to give them a watch and encourage other anglers to watch them, especially those who may need a bit of extra hands-on advice.

Finally, I would like to say a big thank you to Dr Bruce Mann who retired from ORI at the end of February 2023 after more than 30 years of service. Bruce, your leadership and commitment to SAAMBR and the ORI-CFTP has been unwavering, and your dedication to conservation and sustainability of the natural world has truly been inspirational. Thank you for all you have done, and all you will continue to do, as your legacy lives on.

We sincerely hope that you enjoy this online version of the Tagging News. Tight-lines and happy reading!

Acknowledgements:

Financial and administrative support from the South African Association for Marine Biological Research and the KwaZulu-Natal Department of Economic Development, Tourism and Environmental Affairs is gratefully acknowledged.

Thank you to all the anglers who donated funds to the ORI-CFTP in 2022.

We also thank Hallprint[©] Australia for their excellent service and on-going supply of high-quality tags and applicators.

Neels Koekemoer is thanked for his assistance in fitting handles to the tag applicators in 2022.

A special thanks to Marius Els, Derrick Khumalo and Xolani Mselegu for their efforts in capturing and validating the tagging data, as well as for attaching the thousands of tags to tag cards.

ORI-CFTP Fish Measuring Stretcher



The ORI-CFTP is pleased to announce the availability of our purpose made Fish Carrying and Measuring Stretchers.

Made by the competent team at Dive Factory, these stretchers are durable, light, and easy to carry in your fishing bag. They have a measuring tape (150 cm) firmly stuck down the middle of the stretcher with the excess left hanging at the end (for big fish that are longer than the stretcher). They have an aluminium "headboard" used to keep the fish flat and straight which helps improve measuring accuracy. Most importantly these fish stretchers are a perfect tool to help anglers better handle their fish. By carrying and measuring the fish in the stretcher, contact with hot dry surfaces (such as sand, rock or a boat deck) is prevented. For an example of how these stretchers are used you can watch our tagging videos here.

If you haven't done so already, you can purchase a fish measuring stretcher from the ORI Tagging Officer for R150.00 (excl. shipping) by sending your request through to oritag@ori.org.za or by WhatsApping 079 529 0711.





Twenty-one years of surf-zone fish monitoring and tagging in the iSimangaliso Marine Protected Area

By Bruce Mann (ORI Research Associate)



Bruce Mann, Pat Garratt and Simon Chater fishing north of Cape Vidal.

The 21-26 November 2022 were the dates for a bittersweet field trip. This was the last surf-zone fish monitoring and tagging field trip to Cape Vidal. The project started in November 2001 and over the following 21 years we conducted a total of 93 field trips (71 to Cape Vidal, 8 to Sodwana, 7 to Bhanga Nek, 4 to Maphelane and 3 to Mission Rocks). A total of 136 anglers participated on trips, some as part of the core team (to keep fishing effort constant) and others as guest anglers. We spent a whopping 372 days fishing which produced 26 500 angler hours of fishing effort. During this time, we caught a total of 24 681 fish (0.93 fish/angler/hour) from 118 species and 43 families. Of these fish, 11 727 were tagged and released and 1 528 were recaptured (13%), which is double that of the ORI Cooperative Fish Tagging Project.

The primary objectives of this project were: 1) to compare catches (catch rates, species composition and fish size) in no-take areas (where fishing is prohibited) with areas where fishing is allowed; 2) to study movement patterns of key angling species and determine their home range sizes; and 3) to make recommendations regarding improved methods of zonation within the Marine Protected Area (MPA). All these objectives were achieved and the value of no-take areas as an essential conservation tool was

repeatedly highlighted. Some of the main outputs included the following: 1) A PhD degree (BQM); 2) direct inputs into the management and rezoning of the iSimangaliso MPA; 3) contributions to 14 peer-reviewed articles in scientific journals; 4) contributions to two book chapters; 5) 19 presentations at scientific symposia and workshops; 6) numerous public talks to fishing clubs etc.; 7) 11 popular magazine articles; 8) three television documentaries; 9) 93 field trip reports, 21 annual reports and five ORI unpublished reports. I think with this list of outputs the project can undoubtedly be considered a success!

By way of a brief overview, it is interesting to look at some of the science that came out of this project over the years. The first was determining the natural mortality rate of Natal stumpnose in the no-take area for direct input into a perrecruit stock assessment (James et al. 2004). We observed a gradual decline in the catch rate of Natal stumpnose due to the closure of the St Lucia estuarine system (Mann & Pradervand 2007). The largespotted pompano/wave garrick population was assessed as being underexploited but there was evidence of localized overfishing at public access points (Parker et al. 2013). We studied the movement patterns and high residency of the main species recaptured (speckled snapper, cave bass, catface rockcod, yellowbelly rockcod and grey grunter) (Mann et al. 2015). We estimated the optimum size that inshore no-take areas needed to be based on fish movements and home range size (Mann et al. 2016a). The slow growth rate of



Simon Chater and Mike Karon puzzling in big surf.

speckled snapper was determined using tag-recapture data (Mann et al. 2016b). We monitored the recovery of the fish populations in the previously exploited area south of Leven Point using the no-take sanctuary area as a benchmark (Mann et al. 2016c). We investigated the practice of catch-and-release shore angling and whether it is compatible with the conservation goals of MPAs (Mann



The team hard at work at Crayfish Point near Maphelane.

et al. 2018). By collecting fish fin clips, we contributed towards several genetic studies including that of catface rockcod (Coppinger et al. 2019). Excitingly, we contributed towards a study investigating the movement patterns of the iconic giant kingfish using acoustic telemetry (Daly et al. 2019). We studied the movement and growth rate of several species including cave bass (Mann et al. 2020) and giant sand sharks (Jordaan et al. 2021). Surprisingly, the data we collected on movement of speckled snapper was even used in mathematical fishery mobility models looking at harvesting fish outside MPAs (Broadbridge et al. 2022). Finally, we undertook a study looking at the movement of catface rockcod using both dart tagging and acoustic telemetry (Mann et al. 2022).

The achievements of this project are largely thanks to the amazing team of voluntary citizen scientists that enthusiastically participated in this project. I would like to take this opportunity to thank every single one of you for your contributions. It was an incredible privilege to be able to fish in this beautiful area and I sincerely hope that all of you have become ambassadors for our MPAs! When I started this project my bosses at the time scoffed at me and said it was just an excuse to go fishing. I think you will agree, we proved them wrong! The project produced good science, contributed to policy and conservation management and generated support for MPAs amongst recreational anglers – a clear success story.

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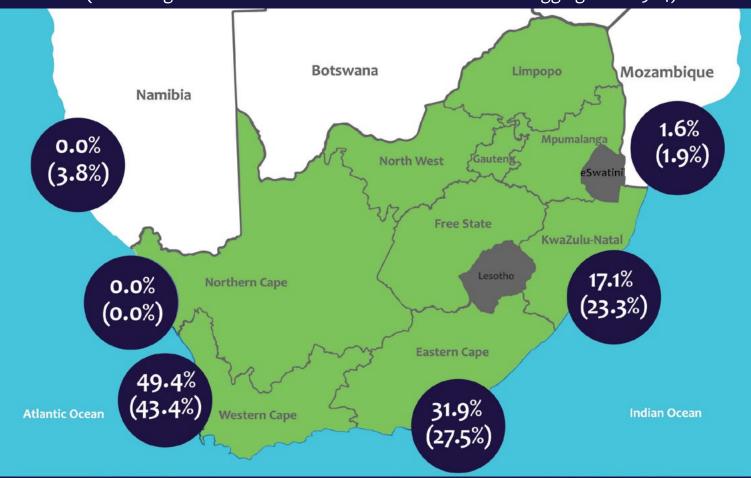
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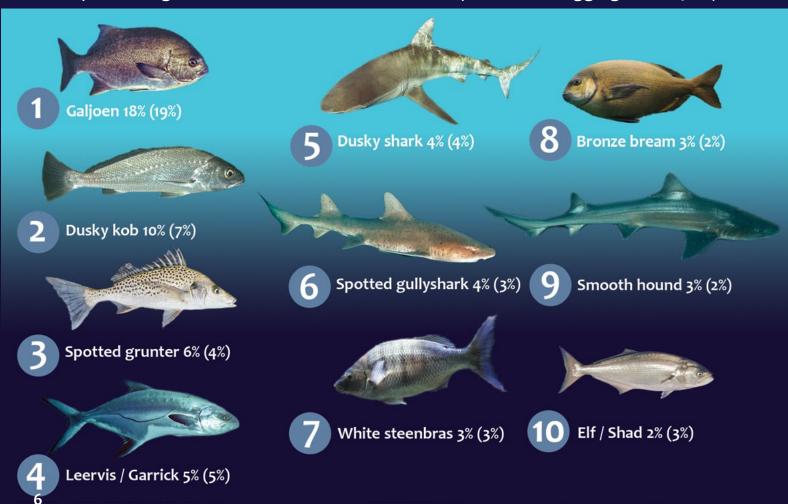
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Percentage of fish tagged along the Southern African coast in 2022 (Percentages in brackets indicate overall distribution of tagging since 1984)



Top 10 species tagged in 2022 (Percentages in brackets indicate overall composition of tagging since 1984)



Get the NEW Fish App for Anglers!

By Bruce Mann

In October 2020, the long-awaited **ORI Fish App** (Marine Fish Guide for Southern Africa) became available for download on cell phone (both Android and iPhone). This app was produced specifically for marine recreational anglers to help improve fish identification and to increase awareness about South Africa's marine linefish species.



All profits from the sale of the App are split between the app developer (PDA Solutions) and ORI. Importantly, funds received by ORI go directly into helping to finance the **ORI-Cooperative Fish Tagging Project** (ORI-CFTP).

The basic structure of the Fish App includes a detailed fish guide (photographs and text), a distribution map for each species, a fish identification tool (smart search), identification guide using fish families, a length/weight calculator, the current fishing regulations for each species and a personal catch log.

The app contains detailed species profiles for **249 common linefish species** from 77 families caught in South African waters, using simple, easy to understand text. The app is very simple and intuitive to use. Excellent colour images for each species have been obtained from a wide range of sources. A useful compare function in the app allows you to compare photos (or text) of similar species. Generalised line drawings of fish families can be used to identify fish in that family. Simple maps are available for the **southern African distribution** of each species. The fish identification smart search is simple to use and works well at narrowing down the species you are looking for. **The length/** weight calculator was compiled



for each species using the most accurate information available and is very quick and easy to use. This is useful when you measure and release your fish but want to know what its weight was. The **linefish regulations have been summarised** for each individual species based on the current gazetted legislation and can be quickly located at the touch of a button. Finally, there is a useful catch log where you can log your own catches and other interesting observations.

Although initial sales have been slow, we hope that the Fish App will become increasingly popular as anglers discover **its usefulness** and spread the word. The app will be regularly updated to include any changes in the fishing regulations and to incorporate any new information on the individual species (updates take place automatically on your phone with no added cost).

To purchase the ORI Fish App, please go to Google Play Store (Android phones) or App Store (iPhones) and search for **"Marine Fish Guide for Southern Africa"**. The app only

costs **R200** to download (less than you spend when you go to the tackle store) so please get yourself a copy now, **enjoy it** and tell others about it!



Research Tagging in Marine Protected Areas

		20	022	Overall		
Marine Protected Areas (MPAs)	Period	Total	# Recapt.	Total	# Recapt.	
De Hoop Marine Protected Area (Western Cape)	1985 - current	1 431	177	64 567	4 900	
Dwesa-Cwebe Marine Protected Area (Eastern Cape)	2009 - current	381	13	5 334	179	
Goukamma Marine Protected Area (Western Cape)	2001 - current	54	2	1 172	40	
iSimangaliso Marine Protected Area (KwaZulu-Natal)	2001 - current	562	40	11 883	1 462	
Helderberg Marine Protected Area (Western Cape)	2021 - current	140	10	582	16	

Top Taggers: 15 or more fish tagged in 2022

Member name	2022 tag releases	Total taggings	2022 tag recaptures	Total tag recaptures	% Recapt.	Member name	2022 tag releases	Total taggings	2022 tag recaptures	Total tag recaptures	% Recapt.
NIC DE KOCK	446	2 637	29	178	7%	WALTER MATHEE	51	351	4	17	5%
MARK GALPIN	268	1 325	24	125	9%	SHAWN METHALAL	49	49	9	9	18%
FRANCOIS VAN ZYL	205	903	10	41	5%	NELIUS SPIES	49	74	1	2	3%
NIKKI-LOUISE SMIT	189	233	4	7	3%	HERMI VAN ZYL	49	76	-	-	-
KEVIN	100	2 6 4 2	-	100	F0/	BOB SHEPHERD	49	829	2	30	4%
	180	2 643	5	122 37	5%	RUAN VAN DER WALT	47	404	2	22	5%
RALDU POTGIETER	175		10	-		ALBERTUS	46	67	3	4	6%
VIVIENNE DAMES	173	445	12	19	4%	NIEUWOUDT					
JEFF ASHER- WOOD	165	912	28	87	10%	GERRIE GROBLER	45	807	7	41	5%
JOHN LUEF	134	949	10	92	10%	GRANGE	45	45	-	-	-
DONAVAN COLE	131	1 274	6	37	3%	MARTIN MALAN	44	49	-	-	-
DIVAN COETZER	127	305	7	14	5%	BRENDAN	43	533	5	82	15%
BRADLEY SPARG	126	2 726	5	158	6%	O'CONNELL					
NIEL MALAN	122	703	7	39	6%	RUSSELL HAND	43	833	-	94	11%
JACQUES DE LA HARPE	119	1 485	6	100	7%	JACQUES-PIERRE GELDENHUYS	41	531	7	46	9%
STEFAN	103	628	15	57	9%	REINER VON DER MARWITZ	41	146	3	8	5%
		4.653			=0/	DYLAN LEES	40	161	3	7	4%
SHAWN MEY	94	1 653	4	80	5%	JAYSON JOOSTE	40	177	1	4	2%
GRAHAM POLLARD	91	497	2	14	3%	WESLEY RAPSON	39	314	-	12	4%
DWAYNE BOSHOFF	87	338	9	17	5%	FRANCOIS KLEYN	39	107	1	6	6%
CHRISTOPHER						ERIC MOREY	39	67	2	3	4%
PIKE	86	345	2	23	7%	STEPHAN OLIVIER	38	147	5	9	6%
BERRIE FERREIRA	82	965	5	34	4%	POENA BRUWER	37	249	-	10	4%
ROLAND NAICKER	77	392	7	23	6%	LLOYD KRIGE	37	57	2	4	7%
BRETT HARRIS	68	307	5	12	4%	SHAUN VAN ZYL	36	419	7	21	5%
CHARLES LILFORD	64	3 399	7	151	4%	FRANCOIS JOHANN VAN DER	36	42	2	2	5%
MATTHEW AND SHANNEN KETHRO	63	67	-	-	-	CHRIS VAN DER					
VICTOR HOGAN	60	160	3	8	5%	WALT	36	75	-	1	1%
	00	100	3	0	376	LYLE TAYLOR	36	390	11	33	8%
MARCO WILDEMANN	59	273	1	6	2%	FRANCOIS KEMP	34	216	7	19	9%
LOUIS LOOCK	59	74	-	-	-	WILLEM SCHOONBEE	33	67	-	3	4%
MARIO ESTERHUIZEN	57	78	-	2	3%	JANNIE VAN BLERK	33	154	3	5	3%
RAY THOMPSON	57	762	4	50	7%	EDUARD STEYLS	33	298	2	9	3%
TARRECK BYRNE	56	139	-	1	1%						
CRAIG NELSON	56	831	1	48	6%	STEPHAN MARX	32	160	2	11	7%
MIKHAIL DANIELS	54	65	6	6	9%	TREMAYNE ANGELO HAMMOND	31	66	7	12	18%
RICHARD MULLER	53	334	2	7	2%	RYAN TAYLOR	31	524	4	51	10%
MATTHEW NOTHARD	52	189	-	-	-	CHRISTIAN JACOBY	30	91	1	6	7%

Top Taggers: 15 or more fish tagged in 2022

	2022 tag	Total	2022 tag	Total tag			2022 tag	Total	2022 tag	Total tag	64 D
Member name	releases	taggings	recaptures	recaptures	% Recapt.	Member name	releases	taggings	recaptures	recaptures	% Recapt.
ROBERT KYLE	29	1 844	3	203	11%	MATTHEW FENN	19	37	1	1	3%
GUY NICHOLSON	28	143	4	8	6%	CORNELIS REIMAN	19	553	-	21	4%
CHENELLE MORAN	27	200	2	12	6%	MATHEW					
JUSTIN MCCARTHY	26	584	1	38	7%	WEEDMAN	19	603	-	83	14%
PIETER DU TOIT	26	262	-	10	4%	STEVE SUTHERLAND	18	51	-	3	6%
MICHAEL PARRIS	26	131	4	9	7%	MATTHEW	18	177	4	18	10%
ENRICO ROBERTS	26	34	-	-	-	MCIVER	10		-	10	10/0
ANTONY	25	49	2	2	4%	CHARL MARAIS	18	852	2	55	6%
SCHEEPERS						EDUAN MOSTERT	18	37	2	2	5%
GEORGE HAY	25	44	-	-	-	ANDRE BRINK	17	58	3	4	7%
JOHN RANCE SNR.	25	409	-	26	6%	JACQUES MALHERBE	17	183	5	15	8%
WET	25	25	-	-	-	LIONEL KORTE	17	79	-	-	-
JJ STRYDOM	25	252	3	15	6%	WAYNE GERBER	17	46	-	-	-
BRADLEY GOUVERIS	23	23	-	-	-	JACO BANNINK	17	68	-	3	4%
STEFAN VAN						JUAN JOOSTE	17	96	-	2	2%
HUYSSTEEN	23	239	-	8	3%	JACOBUS NEL	17	17	-	-	-
NOAH KLOPPER	23	104	4	5	5%	ALAN BRUMMER	16	36	1	6	17%
RUSSEL BERMAN	22	268	-	11	4%	EMILE VAN TONDER	16	33	-	1	3%
SIMON WALKER	22	5 202	3	398	8%	ARTHUR MANN	16	158	1	23	15%
WALDO KLEYN	22	47	-	1	2%	URSULA OTTO	16	166	1	6	4%
ROBERT WELSH	22	51	1	1	2%	CLINTON DUNK	16	83	-	3	4%
BRUCE QUINTIN MANN	22	506	2	43	8%	PHILIP VILJOEN	16	29	1	3 1	3%
YUSUF DHALECH	22	77	8	14	18%	ANDRE FARR	16	62	2	3	5%
PAUL VAN	22	251	4	24	10%	DANIE OTTO	16	30	2	2	7%
NIMWEGEN						EUGENE VAN DER	15	26	_	1	4%
RICHARD COOK	22	151	2	26	17%	ELST					
NIKOS NICOLAIDIS	21	126	2	10	8%	GREGORY MULLER	15	202	-	6	3%
DEAN IMPSON	20	20	-	-	-	RUDOLF TOME	15	17	2	2	12%
CHRISTIAAN ZWIEGELAAR	20	20	-	-	-	MARLIN KINSEY	15	149	-	8	5%
WARREN	20	46	1	1	2%	GARY THOMPSON	15	163	-	4	2%
KNEZOVICH	20	40	-	1	2 %	GOOSEN LE ROUX	15	31	2	4	13%
DEON VAN EMMENIS	20	125	-	4	3%	ANDRE VAN NIEKERK	15	57	2	2	4%
CHARLES DE LA HARPE	20	541	4	64	12%	WILLEM WESSELS	15	40	-	1	3%
GARETH GOUGH	20	637	1	54	8%	-					
JOHN MONG	20	63	1	6	10%	Well done to our top taggers. If you would like to					o view
ROBERT TUZZA	20	36	-	-	-	this year's leaderboard so far, please follow this lin					
DAVID SCHENCK	19	331	2	21	6%	www.oritag.org.za/Leaderboard					

VAUGHN REILLY

TINUS VAN

STADEN

19

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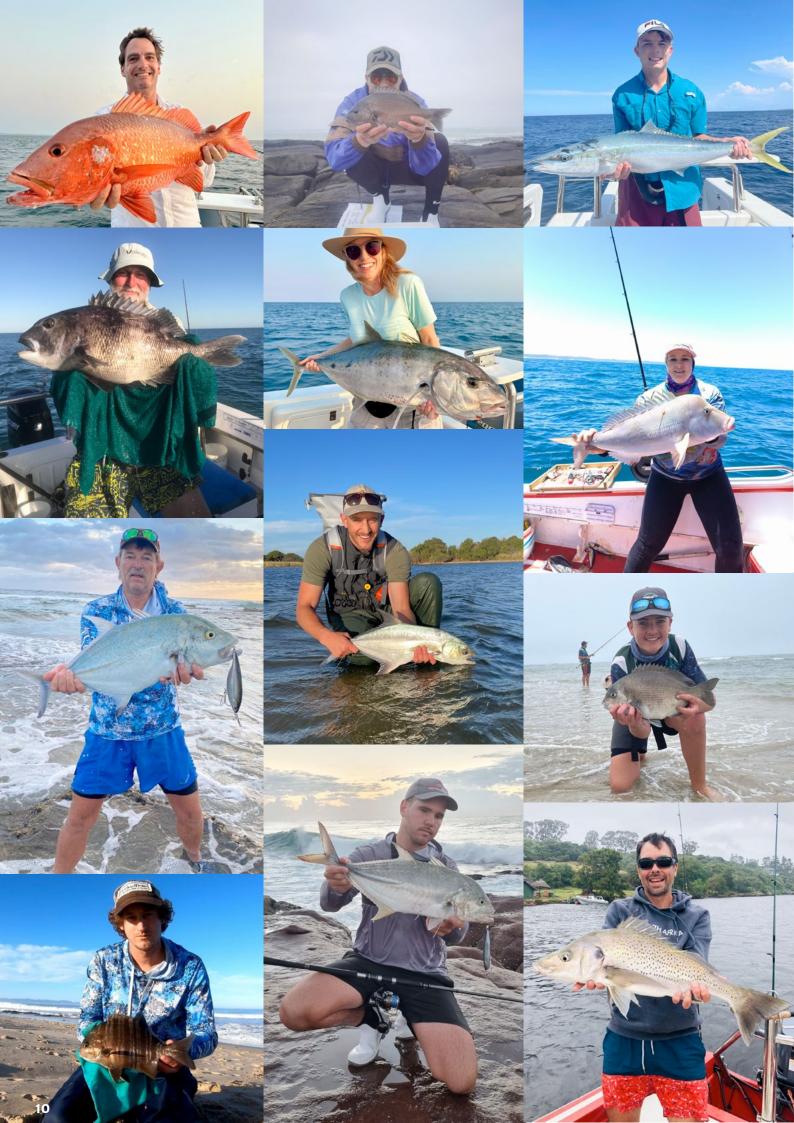
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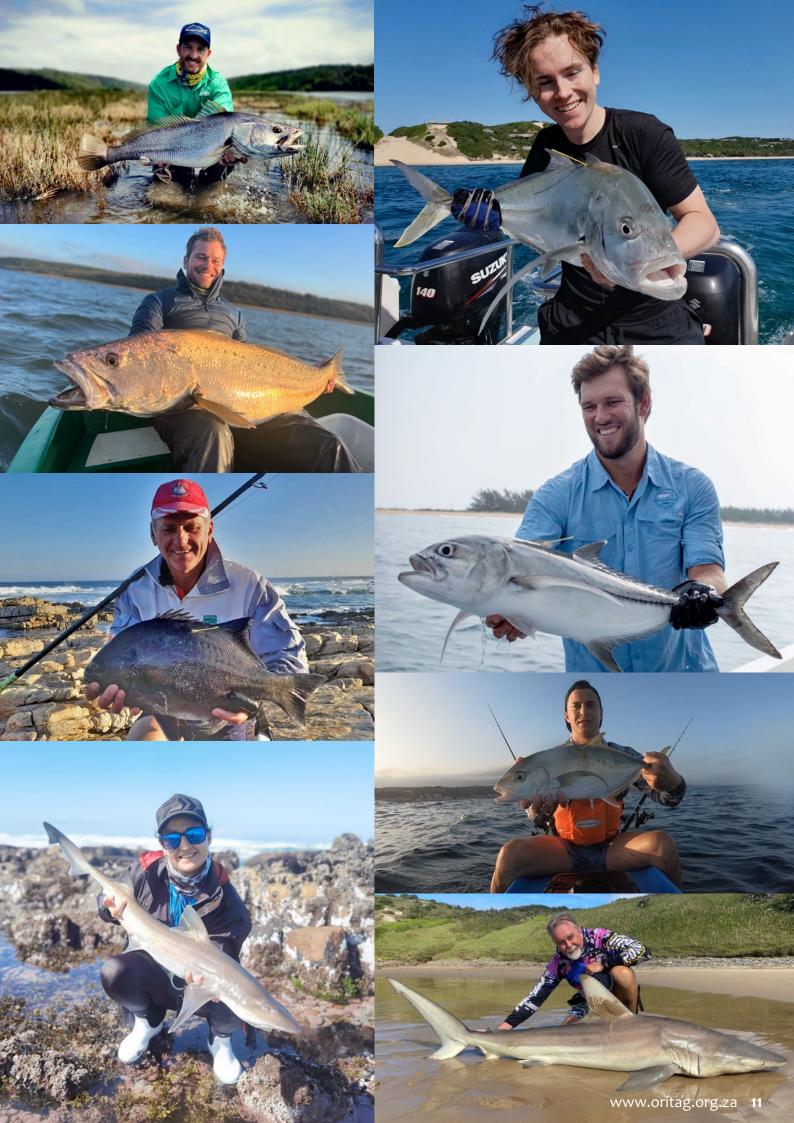
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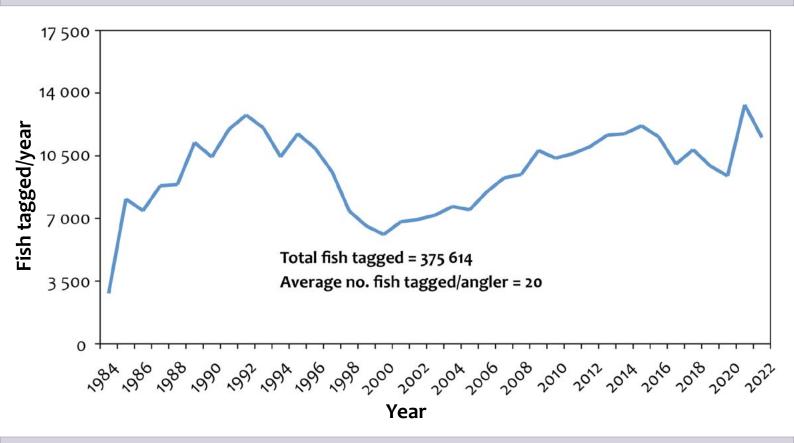




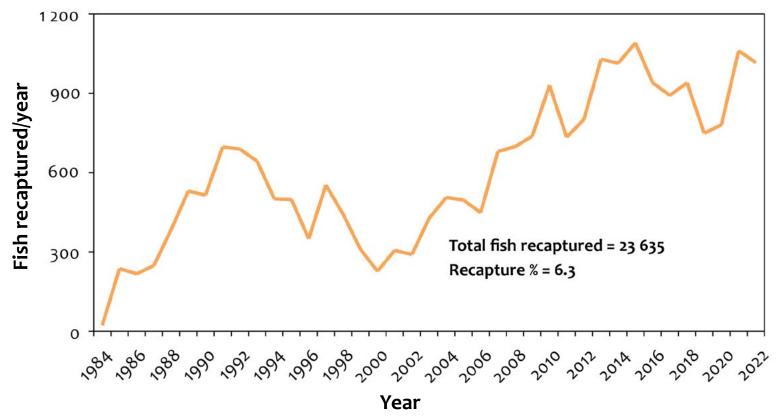


ORI Cooperative Fish Tagging Project Statistics

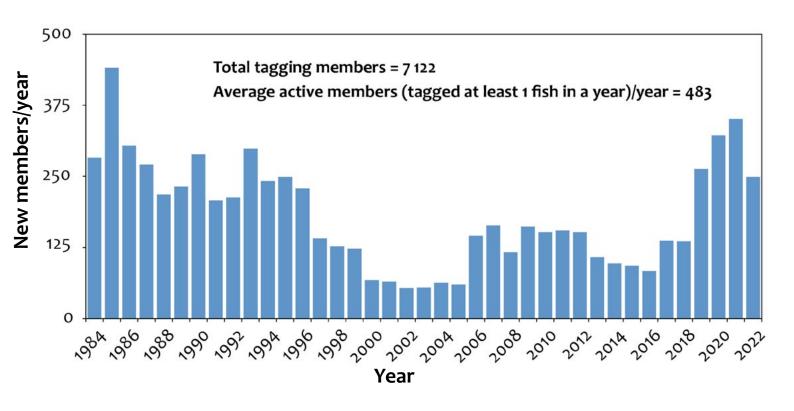
Fish tagged per year and per angler



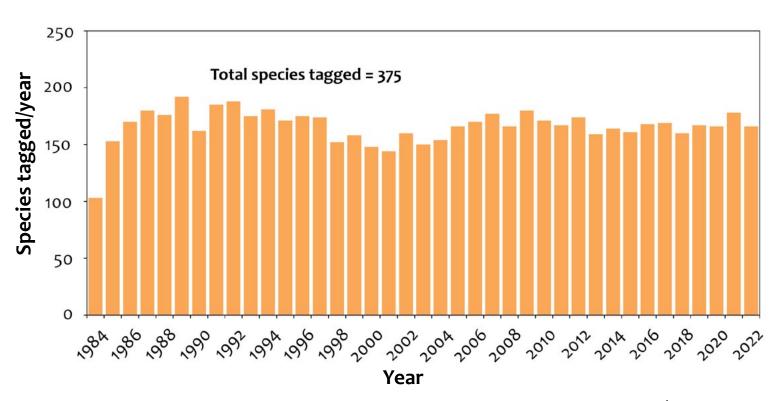
% fish recaptured per year and cumulative number of fish tagged



New members per year



Total species tagged per year



www.oritag.org.za 15

Main fish species tagged up to 31 December 2022

Priority species for tagging are highlighted in blue Recaptured Km travelled Davs free Recaptured Km travelled Davs free No. Tagged No. Tagged Species since 1984 Species since 1984 since 1984 since 1984 No. % Avg. Max. Avg. Max. No. % Avg. Max. Avg. Max. 72 265 5 071 42 1 892 438 7 356 757 286 1 4 4 1 Galjoen 7% Brassy kingfish 1 5 2 5 80 5% 11 Dusky kob 1 813 7% 351 5 997 Dageraad 8% 592 403 25 467 28 1 6 2 5 1 506 119 23 2 3 5 5 Leervis / Garrick 1 4 2 1 7% 216 2 0 6 0 322 3 208 88 6% 21 251 1 292 19 683 1 500 1 Grev grunter King mackerel / 2 950 Spotted grunter 16 747 435 3% 13 823 296 1 4 2 4 61 4% 366 1 5 5 2 534 2 604 Dusky shark / Grey 16 527 1 5 2 6 9% 59 1 374 108 2 9 2 8 1 4 1 0 10 1% 7 56 204 732 Cape stumpnose shark Copper / Bronze whaler shark Westcoast steen-362 167 3 981 1 4 4 9 11 111 3% 1 7 9 0 428 1 311 78 6% 61 280 253 bras 1 427 Spotted gullyshark 10 681 777 7% 31 911 552 6 3 3 2 Duckbill ray 1 2 7 1 14 1% 42 402 648 Soupfin shark / Vaalhaai Elf / Shad 10 107 397 4% 275 1 676 178 1 437 1 0 3 4 3 586 1 2 2 2 31 3% 137 717 White steenbras 9 472 474 5% 34 804 280 2 262 Blacktip shark 1 2 1 6 42 3% 86 1 288 206 1 1 4 8 Blacktail / Dassie 6 2 715 9 361 224 2% 358 279 1 2 1 3 288 24% 2 015 Dark shyshark 3 86 143 Blackspotted 4 405 8 4 3 9 246 3% 42 582 572 Leopard catshark 1 2 1 1 219 18% 8 722 327 4 4 3 1 smoothhound shark Scalloped hammer-Raggedtooth shark 7 3 4 9 1 171 16% 186 2 966 729 9 591 2 943 1 1 7 4 18 2% 121 629 329 head shark Lesser guitarfish / 75 6 6 4 6 1% 43 726 335 2 572 9 Stonebream 1 0 9 7 1% 75 524 242 563 Sandshark Giant guitarfish / 480 8% 32 360 2 8 1 6 5 7 5 9 390 Giant yellowtail 1 0 6 6 45 4% 170 1746 319 1 380 Sandshark Bronze bream 5 6 8 7 174 3% 17 799 190 1 4 6 5 Skipjack tuna 1 0 4 4 2 0% 536 1 0 6 1 1046 1 628 5 599 377 7% 4 294 386 8 134 Roman Yellowfin tuna 1 0 0 9 14 1% 804 5 6 4 5 319 1 314 Slinger 2 8 1 4 5 2 5 3 209 4% 36 1 1 1 0 221 Milkshark 983 26 3% 87 363 181 772 Black musselcracker Geelbek / Cape 4 723 331 791 6 809 7% 30 588 11 1% 105 904 2 569 956 335 Poenskop salmon Largespotted pom-4 5 1 4 78 2% 12 270 246 1 372 2 751 **Bigeve kingfish** 955 39 4% 12 163 246 paño Yellowbelly rockcod 736 425 3 309 4 4 8 1 16% 6 376 9 266 149 2 043 Squaretail kob 950 67 7% Giant kingfish 4 2 0 1 164 4% 15 419 366 2 2 2 6 Honeycomb stingray 916 18 2% 1 8 313 2 543 Diamond / Blacktip kingfish 908 29 3% 4 54 147 545 4 0 9 2 36 1% 165 1 756 431 2 184 Butterfly ray 3 1 382 Black marlin 857 0% 3 633 163 240 Catface rockcod 3 973 946 24% 6 525 171 2 867 Broadnose sevengill Eagleray 808 8 1% 8 49 442 1 582 4 332 3 967 258 7% 64 597 503 Spinner / Longnosed blacktip shark 194 789 27 3% 87 1 0 5 5 1 295 1 2 1 7 Blue stingray 3 6 2 9 13 0% 30 362 234 Seventy-four 758 27 4% 65 521 559 2 845 Zebra / Wildeperd 3 6 2 4 79 2% 2 52 240 1 399 Potato bass 32 5% 2 22 358 2 6 3 9 Sailfish 3 604 29 1% 61 1 0 6 0 150 727 675 White musselcracker Tiger shark 623 29 5% 267 4 0 6 7 379 1 823 97 56 843 588 3 4 9 9 3 2 7 0 3% / brusher Baardman / Belman Hardnosed smooth-609 9 1% 87 340 344 870 4 870 2 908 45 2% 1 17 417 / Tasslefish hound shark Janbruin / John 600 18 3% 2 15 130 502 Speckled snapper 2 801 1 0 0 9 36% 3 200 292 2 662 Browr Carpenter / Silverfish Natal seacatfish 597 233 39% 0 22 378 2 586 24 290 932 4 766 2 6 0 3 1% 46 Bonefish 4 1% 10 34 122 354 179 7% 18 490 239 1 779 567 Santer / Soldier 2 5 1 9 2 0% 805 848 202 379 Striped catshark 2 382 220 9% 6 381 356 2 5 9 7 Striped marlin 564 Red / Copper 18% 214 116 923 9 257 Halfmoon rockcod 560 100 1 49 513 3 1 8 9 2 0 8 1 10% 878 steenbra Bull / Zambezi shark 534 32 6% 76 539 328 2 599 Sharpnose stingray 465 1973 6 0% 6 24 198 Natal stumpnose / Great white shark 523 17 3% 290 1 5 4 3 346 959 230 1 451 1864 53 3% 14 233 Yellowfin bream Smooth hammer-10% 8 102 519 50 222 997 Brown shyshark 1 851 22 1% 133 384 555 3 075 head shark Queen mackerel / 465 3 1% 4 12 376 1 0 4 4 Ladyfish / Springer 1 4 2 6 1823 36 2% 21 412 376 Natal snoek Blue marlin 454 0 0% 0 0 0 0 4% Silver kob 1715 72 44 548 290 1 4 3 5 894 11 107 1 998 Red stumpnose 446 2% 11 Perch / River bream 238 105 352 1 583 1 6 9 1 14% 1 Cavebass / Lampfish 1 6 5 7 250 15% 9 514 352 3 116 Southern pompano 442 26 6% 62 464 151 848

River snapper / Rock

Albacore / Longfin

salmon

tuna

Scotsman

292

414

36

18%

26%

2%

1 5 8 1

1 572

1 569

3

26

304

391

1 211

1 008

2 403

2 839

2 585

Puffadder shyshark

Lemonfish

Hottentot

327

465

412

441

427

415

41

17

16

9%

4%

4%

1

4

1

20

64

10

234

230

251

1 363

749

1 078

Main fish species tagged up to 31 December 2022

Priority species for tagging are highlighted in blue Km travelled Recaptured Days free Recaptured Km travelled Days free No. Tagged No. Tagged Species since 1984 Species since 1984 since 1984 since 1984 Avg. Max. Avg. Max. No. % Avg. Max. Avg. Max. No. % Talang / Largemouth 4% Maasbanker 0% queenfish Pickhandle bar-14% 1 856 Longfin kingfish 1% racuda 5% White stumpnose 1% Bigeye stumpnose Bluefin kingfish 4% Oxeye tarpon 0% Flapnose hound-Spotted spiny 14% 3 013 1% dogfish shark 1 263 1 263 Swordfish 1% Bartail flathead 3% 1 947 Banded catshark 11% 1 155 Banded galioen 2% Sandbar shark 2% Java shark 3% Eastern little tuna / З Round ribbontailray 4% 0% Kawakawa 0% Blue kingfish Blackspot shark 3% Striped mullet 2% 4% Spearnose skate St. Joseph / Ele-phant fish Minstrel rubberlip 3% 0% 1 342 1 3 4 2 7% Sand steenbras 3% Blue emperor Bluntnose spiny 1% 1 476 Cape moony 0% dogfish Sailfin rubberlip 0% Snapper kob 4% Blue hottentot 3% Dusky rubberlip 4% 1 4 9 5 2 345 Doublespotted Malabar rockcod 15% 1 540 0% queenfish Needlescaled 2% Englishman 4% queenfish False thornback 4% Green jobfish 3% skate Whitespotted smoothhound shark Spadefish 2% 2 724 2 724 2% 1 627 Prodigal son / Cobia 2% 1 895 White seacatfish 2% 0% Yellowtail scad Greyspot guitarfish 1% / Sandshark 10% Shortfin mako shark Snoek 1% Marbled electric ray 0% Shorttail stingray 3% 2 4 1 2 Thintail thresher 0% shark 2 940 9% Javelin grunter Concertina-fish 0% Dorado / Dolphinfish 1% German 0% Spotted eagleray 2% Swallowtail rockcod 9% Striped threadfin 1% Panga 0% 2 990 Moustache rockcod 29% 1 200 Yellowfin emperor 9% 1 187 16% Tomato rockcod Shortbill spearfish 0% Smallspotted pom-3% 1 176 1 176 Koester 2% pano Greater yellowtail / Amberjack Wreckfish 5% 2% Bludger kingfish 0% Grey reef shark 2% Blue shark 0% Yellowspotted 0% Captain Fine / Whitespotted kingfish 0% Cock grunter 4% rockcoc 0% Indian goatfish Thorntail stingray 2% 0% Steentije Longfin / Tropical 3% yellowtail Manta 3% 21% Great barracuda Threadfin mirrorfish 0% Whitebarred 1% Surge wrasse 3% rubberlip Russell's snapper 3% Tripletail 0% 0% Atlantic bonito Twinspot snapper 16% Flathead mullet 1% Wahoo 3% Cape gurnard 3% Indian mirrorfish 0% Eel catfish 1% Milkfish 0%

1 3 3 4

2 6 5 2

Mackerel

0%

2%

Sliteye shark

www.oritag.org.za 17

Port of Ngqura: An unexpected, artificial, sanctuary for fish.

Written by: Vivienne Dames, Matt Dicken and Tony Booth

The Port of Ngqura, called Coegha Harbour, is 20 km east of Gqeberha, South Africa. The deep waters within this port were developed by extensively dredging the mouth of the small and temporary Coegha River. Construction was completed in April 2006, and the port became operational at the end of 2009. The structure was an impressive feat of engineering that has turned

red tides. During upwelling events in Algoa Bay, it is likely that many species, and their juveniles, will seek refuge in Port Ngqura's slightly warmer temperatures. These warmer temperatures also support a unique mix of cool-temperate and tropical species. To date, incredible sightings have been made of aggregating smoothhound sharks, gulley sharks, ladyfish/springer



Figure 1: The port of Ngqura, which began commercial ship operations (containers) in October 2009, is situated 20 km northeast of Gqeberha and is South Africa's 8th and latest commercial port development, situated at the mouth of the small ephemeral Coega River in Nelson Mandela Bay (Algoa Bay). Image: Oceanic Seagull Maritime

what was once a tiny estuary between expanses of beach into a harbour with an industrial development zone, with one breakwater 1.3 km and the other 2.7 km long. These breakwaters were constructed from rock armoury and 26 500 concrete dolosse weighing 30 tonnes each. Several microhabitats exist in the port, including dolosse, rock armoury, shallow profile reefs, vertical quay walls and the small sandy beach, which still exists at the mouth of the Coegha River.

The incredible diversity of species studied within Port Ngqura bears no resemblance to the sea life around the beach that was dredged in 2006 – and heartening to see a new suite of marine characters making lives for themselves in a man-made environment. These species are suggested to utilise the calm, deep waters of Port Ngqura, where adjacent shores are typically rough. Species, such as raggedtooth sharks, have been observed to increase in abundance during unfavourable and garrick/leervis. Other sightings include whale sharks, bull sharks, manta rays and even great whites when there has been a whale carcass in the area. It is also not uncommon to see large pods of dolphins in excess of 200 individuals and even Bryde's whales enter the port.

In 2006 a small group of dedicated anglers spearheaded by Prof Matt Dicken established the Port Ngqura long-term biomonitoring programme. This biomonitoring programme involved Bayworld Research, KZN Sharks Board, the Oceanographic Research Institute's Cooperative Fish Tagging Project (ORI-CFTP) and the Transnet Environmental Management

team based at Port Ngqura. This project aimed to document the species composition and relative abundance of fish within the port, investigate the size composition of fish within the port and the function of the port as a nursery area for juvenile fish, and identify



Figure 2: An aggregation of spotted gulley sharks (Triakis megalopterus) along the small beach within Port Ngqura. Similar aggregations of smoothhound sharks (Mustelus mustelus) are more commonly seen, in greater numbers every summer. Image: Prof Matt Dicken.

spatial patterns in the distribution of fish associated with different habitats within the port. It is important to note that public access to fishing is prohibited within the breakwaters of Port Nggura.

Several years later, long-term dart tagging and monitoring has produced a massive dataset and shown the incredible biodiversity of fishes in this artificial man-made seascape, with exceptionally high densities of sharks and rays. Catches by trained teams of anglers were recorded on standardised datasheets between September 2006 and September 2007 and from December 2011 to the present, to monitor the health of the Port environment. Best handling practices (use of buckets, mats and limiting air exposure) and 100% catch-and-release are mandatory in the biomonitoring programme. The long-term biomonitoring programme is ongoing and comprises a team of 15 anglers participating all year round. Between September 2006 and September 2007, 4 559 fish were caught. From December 2011 to the present, an additional 12 643 fish (comprising 72 different species) were caught and released, of which 7 507 have been tagged with spaghetti tags via the ORI-CFTP. From dart tagging, 698 (10%) fish have been recaptured. Tagged specimens comprised 62 different species, with the dusky shark being the most commonly tagged (n = 1 451) and recaptured species (11.7%), followed by garrick with 1 277 tag releases and 8.8% recaptures. Interestingly just over half of all recaptures in Port Ngqura have been reported by members of the biomonitoring programme, indicating a strong residency of some species (e.g. dusky kob, santer, yellowbelly rockcod) and the return of mobile species (e.g. dusky shark, garrick, ladyfish).

Regarding the movement patterns of all species combined, around 7% moved within a small range of 6-10 km, 4% moved 11-20 km, and 3% moved 21-50 km. The furthest movements were accounted for by 6% of recaptures, having moved between 501-1000 km. The farthest recorded movement was 950 km, achieved by a garrick originally tagged at the Port of Nggura on June 23 2018, measuring 790 mm in fork length (FL). This garrick was recaptured 377 days later at St Lucia Beach, KwaZulu-Natal (KZN), measuring 850 mm FL. Notably, the top three movements were all made by garrick, with an average distance of 261 km. These recaptures support previous research showing that garrick is a highly migratory species. It also implies that the Port of Ngqura (along with the surrounding estuaries) may serve as an important area for juvenile garrick before they embark on their seasonal spawning migrations to KZN as adults (for more details, refer to Dunlop et al. 2015). Other species exhibiting significant movements include raggedtooth sharks (max movement of



Figure 3: A close up of a raggedtooth shark (Carcharias taurus) in clear visibility. Clear visibility is common in the Port environment during summer when water temperature ranges between 22 and 25°C. This picture also shows the established reefs found along the breakwalls of Port Ngqura. Image: Vivienne Dames.

892 km), elf/shad (max movement of 768 km), dusky sharks (max movement of 731 km), and blacktip sharks (max movement of 455 km), all of which are known to be migratory species.

Analysis of the 2006-2007 dataset has produced two scientific publications. One was primarily focused on dusky sharks and showed the Port was a critical summer habitat for juveniles, with population size estimates of approximately 552 juveniles in the port during the summer months. Catch per unit effort (CPUE) was greatest between October and February, peaking in November at 0.51 sharks/angler/hour (see Dicken 2011 for more information). The 2006-2007 dataset had at the time recorded 52 species. A second publication analysed habitat use and catch composition between shore and boat angling. Shore-angling was shown to have a CPUE of 2.3 fish/angler/h and boat-based angling 2.8 fish/angler/h. Catches included species representative of both estuarine and shore fisheries. Catch composition differed significantly between the dolosse, quay wall and sandy shore habitats. Of these three micro-habitats, the dolosse were shown to support the greatest abundance and diversity (See Dicken 2010 for more information).

The biomonitoring programme has recently branched out to include a study using underwater video surveys. The surveys are conducted using baited-remote underwater video systems (BRUVs). Two cameras are mounted in a metal frame, with an arm extending forward and holding a bait cannister containing crushed sardine. These systems are then deployed overboard for one hour before being retrieved. The systems are deployed within all the Port habitats, at various depths and throughout the year. This research aims to provide another method for collecting data on fish species diversity, abundance and size data while also looking at



Figure 4: A close up of a resident yellowbelly rockcod (Epinephelus marginatus) on an established reef in winter, when the water in Port Ngqura has lower visibility. Image: Vivienne Dames.

habitat complexity. It will be interesting to see how the fish communities recorded on the BRUVs differ from those in the catch-and-release portion of the project. So far, 120 BRUV deployments have been completed since August 2022.

Although not formally analysed, this footage shows numerous, fully established reef ecosystems along the breakwalls. These reefs are covered in sponges, bryozoans, algae and are teaming with fish life. With these videos, we have discovered a diverse array of small tropical species utilising the Port in the summer months as a unique ecosystem. Several species which catch data has yet to document have also been added to the extensive species list in Port Ngqura. Added species include roman, dageraad, black and white musselcracker, galjoen and white stumpnose populations. In summer, high abundances of smoothhound, dusky sharks, raggedtooth sharks and diamond rays frequent these camera systems. It is without a doubt that this new angle of research will



Figure 5: Underwater picture of a tagged garrick (Lichia amia) in Port Ngqura, a popular recreationally targeted species. This is the second most caught, tagged and released fish in Port Ngqura with 1 842 caught, 1 277 tagged and 8.8% recaptures. Image: Vivienne Dames.

greatly contribute to our understanding of how fish communities utilise Port Ngqura, what their reasons are and what role the artificial reef structure plays in forming these unique communities. We would like to make special mention of the Save Our Seas Foundation (SOSF) and the South African Institute for Aquatic Biodiversity (SAIAB) for making this exciting research possible.

Although the port's primary objective is not as a recreational fishery, we hope that management will attempt to retain the biological potential of the Port by using design and construction features that provide enhanced fish habitat. Retention of the nursery function of the Port to many juvenile fish and shark species is essential considering the continued degradation of



Figure 6: An adult scalloped hammerhead (Sphyrna lewini), a critically endangered species. Aggregations are commonly seen in Port Nagura during the peak summer months, mostly consisting of juveniles. This species is handled with particular care, not being taken out of the water and released as guickly as possible. Image: Vivienne Dames.

many estuaries, which are vital in the life histories of so many overexploited linefish species. The diversity and abundance of fish recorded within the Port of Nggura suggests the potential use of ports and other artificial structures for enhancing the conservation potential of man-made seascapes for local fisheries in South Africa. We build defences against the ocean's power to keep our cities safe and dig out harbours to facilitate trade. In some developed countries, such as the UAE for example, more than half the coastline is artificial. To ensure that we keep these existing manmade structures fish friendly, we need to find out how animals adapt to them and shape their lives around them. In a man-made marine environment, this could prove crucial to their conservation.



Exciting Recaptures From 2022



Total distance moved = ±1 308 km Total growth = 40 mm Total time at liberty = 208 days

On the 26th October 2022 the ORI Cooperative Fish Tagging Project (ORI-CFTP) had its 399th elf/shad tag recapture! This fish was originally tagged by a team of citizen scientists at Lekkerwater, Western Cape (WC), during a field trip in the De Hoop Marine Protected Area (MPA), on the 21st February 2022, measuring 600 mm total length (TL). It was recaptured by Amith Kanthapersad having travelled an incredible 1 308 km north at Amanzimtoti / Nyoni Rocks, KwaZulu-Natal (KZN), just 208 days later, measuring 640 mm TL. It was recaptured during the closed season, so it was released again, hopefully to contribute to spawning that takes place at this time of year. This species is considered to be overexploited in South African waters and because of their compromised stock status, there is a strict daily bag limit of 4-per-person-per-day, a minimum size limit of 30 cm TL, and a closed season from the 1 October until the last day of November each year.



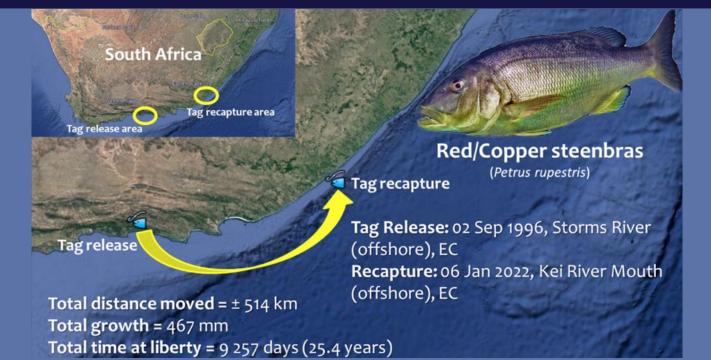
On the 23rd February 2022 we had our 38th soupfin shark (vaalhaai) tag recapture for the ORI Cooperative Fish Tagging Project (ORI-CFTP)! This soupfin shark was originally tagged by Jan Pieterse on the 14th December 2019 in Stilbaai, Western Cape (WC); unfortunately no measurement was taken. This shark was recaptured 792 days (2.2 years) later by Chris Burley in Summerstrand near the Something Good restaurant, Eastern Cape, measuring 112 cm pre-caudal length and having moved about 489 km up the coast. Unfortunately, no growth rate could be derived from this recapture. A recent stock assessment from 2019 indicated that this species is severely overexploited in South African waters and that current commercial catches are not sustainable. They have been evaluated as Critically Endangered on the IUCN Red List (2020).



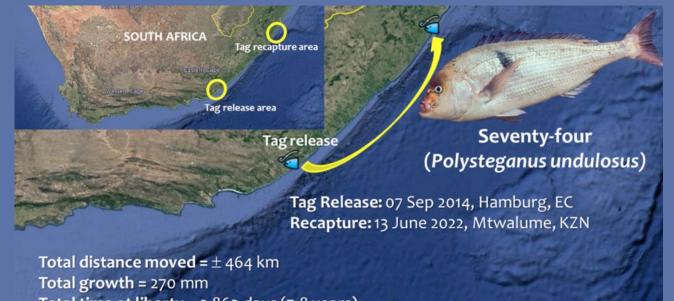
On the 26th August 2022 the ORI Cooperative Fish Tagging Project (ORI-CFTP) had its 166th giant kingfish (also known as GT or giant trevally) tag recapture! This fish was originally tagged by Peter Stewart whilst out on a charter trip with Mozambique Angling Adventures on the 27th June 2019 at a reef near Ponta Abril (Santa Maria/Hells Gate), Mozambique, measuring 900 mm fork length (FL). It was recaptured by Barry De Beer around the same reef in Ponta Abril 1 156 days (3.2 years) later. Unfortunately, no measurement was taken, but it was estimated to be about 105 cm and was released again to fight another day. Giant kingfish receive protection from capture in several no-take Marine Protected Areas (MPAs) along the east coast of southern Africa such as within the Maputo, iSimangaliso, Aliwal Shoal and Pondoland MPAs. Particularly sensitive areas include the Mtentu Estuary within the Pondoland MPA and an area within the Maputo National Park MPA where seasonal spawning aggregations take place.



On the 11th February 2022 we had only our 15th duckbill ray tag recapture for the ORI Cooperative Fish Tagging Project (ORI-CFTP)! This ray was originally tagged by Hansie Pretorius on the 3rd January 2020 at Bluewater Bay/Swartkops, Eastern Cape, measuring 117 cm disc width (DW). It was recaptured by Wian Bloem 770 days (2.2 years) later at the Klein Brak River, Western Cape, having moved about 402 km west, now measuring 128 cm DW having grown 11 cm. Duckbill rays are mainly found inshore down to depths of 150 m. They are sometimes seen at the surface leaping out of the water to rid themselves of parasites or suckerfish/remoras. They are extremely powerful fish and well respected by shore anglers. There is limited tag recapture data from this species, but what we have seen is that this ray is relatively mobile with an average distance moved of 41.2 km and a maximum distance moved of 402 km. A study using acoustic telemetry is now being conducted on this species.



On the 6th January 2022 we had a red steenbras tag recapture, which turned out to be a record teleost (bony fish) recapture for the ORI-CFTP! This fish was originally tagged at Middlebank off Storms River, EC, on the 2nd September 1996 by Dr Steve Brouwer (then with Rhodes University) during a research tagging field trip in the Tsitsikamma National Park. The fish originally measured 563 mm FL. It was recaptured an astonishing 25.4 years later offshore of the Kei River Mouth by Gary Thompson (with the same tag still intact - thanks to our ever-reliable tag suppliers, Hallprint©, Australia), measuring 1030 mm FL, and having moved about 514 km. According to David Hall, Managing Director of Hallprint, this is the second longest record time at liberty for a teleost fish amongst all the tagging programs that they supply tags to. The record is currently held by a southern bluefin tuna tagged off southern Australia that was at liberty for 9 639 days (26.4 years)!



Total time at liberty = 2 863 days (7.8 years) he 13th June 2022 we had our 26th seventy-four tag recapture for the OF

On the 13th June 2022 we had our 26th seventy-four tag recapture for the ORI-CFTP, and our first one since 2017! This seventy-four was originally tagged by Brendan Croney on 7th September 2014 at Keiskama Point/Hamburg, EC measuring 500 mm FL. It was recaptured 2 863 days (7.8 years) later by Alan Fraser off Mtwalume, KZN measuring 770 mm FL. This individual moved about 464 km north and grew 270 mm during its time at liberty. The fish was re-released again hopefully to be able to spawn and contribute to population recovery. The Dwesa-Cwebe, Amathole and Addo MPAs likely play an important role in providing protection for juvenile seventy-four. The Pondoland, Protea Banks, Aliwal Shoal and Thukela MPAs will likely play a role in the protection of adults. In particular, the northern extension of the Aliwal Shoal MPA to include part of the Illovo Banks (which was one of the historic spawning grounds of seventy-four) will hopefully assist in protecting spawning aggregations of this species and help to ensure its recovery.

[Acoustic] Tag, you're it!

By: Taryn Murray

The Oceanographic Research Institute's Cooperative Fish Tagging Project (ORI-CFTP) is one of a few citizen science programmes worldwide worth boasting about. Not only has it been running for almost 40 years, which in itself is



Duckbill ray on PVC sheet - Matt Parkinson

incredible, but the information collected by the ORI-CFTP over the past (almost) 4 decades has proven absolutely invaluable, allowing researchers to learn so much about the movements of multiple fishes, sharks and (to a lesser extent) rays.

Another method, whose data can complement the broadscale nature of the ORI-CFTP data, is acoustic telemetry. This method essentially makes use of two pieces of equipment, which work via sound:

1) an acoustic receiver which is deployed in rivers, estuaries or at sea, and

2) an acoustic transmitter or tag, which is externally attached onto or surgically inserted into an animal, and can have a battery life of up to 10 years.

By placing the receivers in strategic locations, researchers



Receiver with floats in the sea – Ryan Daly

can use the fine-scale high resolution data collected by the receivers to learn more about how much time tagged animals spend in certain places (residency), whether they return to certain areas (site fidelity), whether they undertake annual migrations, and so much more.

Acoustic telemetry is currently the most popular method globally to study the movements of aquatic animals. This popularity has led to the development of several largescale acoustic receiver networks, spanning hundreds to thousands of kilometres, and situated in multiple different countries around the world. These include Australia's Integrated Marine Observing System's Animal Tracking Facility, the European Tracking Network, the global Canadian-based Ocean Tracking Network (OTN), and South Africa's very own Acoustic Tracking Array Platform (ATAP). This acoustic receiver array, which was formalised in 2011 and is currently made up of 300+ receivers, covers ~2200 km of the South African coastline, from False Bay in the Western Cape, to Ponta do Ouro at the South Africa-Mozambique border. The entire network is collaborative



Mtentu receiver – Ryan Daly

in nature, meaning that receivers belong to many different organisations including universities, research institutions and non-government organisations, but all the data collected on these receivers are placed into ATAP's central database, and is freely available to the various tag owners. This is a win-win situation for researchers in that they benefit from the nationwide infrastructure, and it opens doors for potential collaboration.

While the ATAP is now considered a 'mature' network, it had humble beginnings with a single study that started in the West Kleinemonde Estuary in the Eastern Cape, where one large dusky kob, affectionately named 'Walter', was tagged and manually tracked for a week in 2002 (one fish, one species, one estuary). This then led to work on dusky kob and spotted grunter in the Great Fish Estuary between 2003 and 2004 (many fish, two species, one estuary).



Catface surgery – Bruce Mann

Eventually this work was expanded to include other estuary-dependent (and important fishery) species such as white steenbras, leervis/garrick and Cape stumpnose, and receivers were placed in a number of Eastern Cape estuaries (many fish, many species, many estuaries). A partnership was then struck (and formalised) with the OTN, which saw the ATAP receive a loan of 100 acoustic receivers in 2011. These receivers were initially placed in large coastal bays such as False Bay, Mossel Bay and Algoa Bay, but as more equipment was bought, other important coastal regions, such as Gansbaai, Port Alfred, Port St Johns and Sodwana Bay, received acoustic coverage too. And so the ATAP came to be, and has steadily expanded since, with many researchers incorporating their localised receiver arrays into the greater network (e.g. Shark Spotters' array in False Bay, Western Cape, and the Oceanographic Research Institute's array along the Wild Coast and KwaZulu-Natal coastlines).

The ATAP currently monitors the movements of 800+ individual animals comprising 38 species. These include important fishery species such as dusky kob and spotted grunter, large predatory sharks such as bull sharks and bronze whalers, Critically Endangered species such as whitespotted wedgefish (formerly giant guitarfish) and the common eagle ray, commercially important sharks such as soupfin and smoothhound sharks, prized sport fish such as giant kingfish and leervis/garrick, small endemic species such as flapnose houndshark and blue stingray, and rehabilitated turtles including green, hawksbill and loggerhead turtles. Overall, the ATAP database has more than 25 million detections, meaning that tagged animals have been recorded moving past a deployed receiver that many times.

When animals are dart tagged (such as volunteer anglers do in the ORI-CFTP), and hopefully recaptured further down the line, there are only two data points for that animal – time and location of capture and tagging, and time and location of recapture. This allows researchers to get a better idea of species' distributions, broad-scale movement information and growth rates. While the usefulness of dart tagging data such as that collected by the ORI-CFTP cannot be doubted, acoustic telemetry data have provided greater insights into the movements of fishes, sharks and rays, and more recently, turtles. This is mostly due to the larger volume of data collected per animal, as well as on a (much) finer scale. For example, the ORI-CFTP rarely reports on fish tagged and recaptured in estuaries, but rather groups these records into one of the designated ORI-CFTP localities along the coastline. In contrast, a telemetry study conducted in an estuary can provide information on hourly, daily and seasonal movements. These movements can also be linked to changes in water temperature or tide, time of day or moon phase. Telemetry has also significantly improved our understanding of the importance of estuaries to juveniles and even adults of important fishery species, such as dusky kob, spotted grunter, white steenbras and leervis/garrick. Another added bonus of acoustic telemetry is gaining a better idea of how much protection marine protected areas (MPAs), in which fishing is generally not allowed, afford different species. This is because receivers can be positioned within MPAs, and on putting the pieces of the movement puzzle together, we can get a better idea of how connected these places are, and how much time certain species might be spending in these areas relative to unprotected areas.



Grey reef shark release - Ryan Daly

Another added, yet unexpected and sometimes scary, benefit of acoustic telemetry is learning more about the fishing mortality of tagged animals. As far as we are aware, most tagged fish that have been recaptured have been reported due to a REWARD sticker being stuck onto the transmitter prior to tagging an animal. For example, of 10 juvenile dusky kob tagged in the Breede Estuary, nine, or 90% have been caught and kept. At least 100 leervis/garrick have been tagged during the past decade, and of these, 21% have been caught and kept, including a recent recapture of a fish eight years later that was tagged in 2015 at a length of 90 cm. But, thanks to acoustic telemetry, we know that this fish undertook its annual spawning migration to KZN every



Green turtle tagged with an acoustic and satellite tag – Linda Ness

single year since tagging – something we would never have known with dart tagging.

Ultimately, each method provides a huge amount of movement information about numerous different species, but when combining both dart tagging and acoustic telemetry, that is when we really begin to learn and understand so much more.

For more information on the ATAP, the research we do, and some exciting movements, be sure to follow us on social media!

Facebook: Twitter: Instagram: ATAP – Tracking fish movements @ATAP_ZA @atap_za



How to report the recapture of a tagged fish

This video below provides all the information that you need to correctly report tag recapture information. Tag recaptures are one of the most important and exciting aspects of the Oceanographic Research Institute's Cooperative Fish Tagging Project (ORI-CFTP). Recaptured fish allow us to investigate movement patterns, growth rates and population dynamics of the fish species tagged along the southern African coastline and ultimately contribute towards their conservation. What makes the ORI-CFTP so interesting and exciting is seeing where a recaptured fish was originally tagged; how far it has travelled; who originally tagged it and how much it has grown. As anyone who is fishing in the sea stands a chance of catching a tagged fish, it is very important to know exactly what information to record and how to send it to ORI.



Focus species Catface rockcod (Mycteroperca [Epinephelus] andersoni)

Movement:	A recent study has identified that this species is likely to be a temporary resident on shallow inshore reefs. Catface rockcod show high residency and occupy relatively small home ranges (a few 100 m ² in extent) for short periods often less than 12 months, whereafter they abandon their home ranges and make ranging-type movements in search of new habitat. Adults >400 mm are more mobile than juveniles, with distance moved increasing with fish size. There are anecdotal reports that they form spawning aggregations, suggesting that they may migrate for this purpose.
Total number tagged:	3 973
Number recaptured:	946 (24%)
Longest time free:	2 867 days or 7.9 years (1992 – 2000)
Longest distance moved:	525 km (Bats Cave [Mission Rocks, KZN] to Praia do Xai Xai, Mozambique)
Growth:	They mature at 43 – 49 cm total length (TL) at an age of 3 – 4 years. Generally, males mature earlier than females, and some females may change sex to male.
Max size:	87 cm TL; 9 kg
Max age:	11 years
Breeding season:	Spring to Summer (September to February)
Breeding location:	In the northern parts of their distribution. There is little evidence of spawning south of Durban.
Feeding:	They feed on crustaceans, small fish and squid.
Distribution:	Endemic to the eastern seaboard of southern Africa, found from Quissico in southern Mozambique to Knysna in the Western Cape. However, some individuals have recently been caught as far south as the De Hoop Marine Protected Area.
IUCN Red List status:	Near Threatened (2018)
SASSI List:	Orange (think twice)
Description in the second	

Recent Publications:

Mann BQ, Daly R, Jordaan GL, Dalton WN, Fennessy ST. 2022. Movement behaviour of catface rockcod *Mycteroperca* (*Epinephelus*) *andersoni* (Epinephelidae) off the eastern seaboard of southern Africa. *African Journal of Marine Science* 44(2): 125-137.

https://doi.org/10.2989/1814232X.2022.2064548

Francois Johann van der Merwe with a 44 cm TL catface rockcod that he tagged and recaptured six days later off Umdloti, KZN.

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Helping people to care for our ocean